

Curvularia leaf spot on white fonio accessions in the commune of Boukoumbe in north Benin

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1 ABSTRACT

Fonio cultivation in West Africa relies only on traditional accessions, which are less productive, and their resistance to Curvularia leaf spot, one of the major constraints, are not well documented. The objectives of this study were to determine the distribution of the leaf spot of white fonio (*Digitaria exilis*) in the commune of Boukoumbe in north Benin, to identify its causal agent and to evaluate white fonio accessions for their resistance to the leaf spot. A survey was conducted in July 2018 in the 6 districts of the commune of Boukoumbe. Two to three sites were surveyed per district and at each site, 30 plants were randomly selected on two diagonals through the field to evaluate the disease incidence. Ten of these plants were used to assess the disease severity. Samples of the diseased plants were taken to the laboratory for isolation. Additionally, an experiment was conducted in a greenhouse at the experimental farm of the faculty of agronomy of the University of Parakou (Benin) where twenty-three accessions were inoculated with two virulent isolates of the pathogen, and evaluated for disease severity and seed weight losses due to the disease. The experiment followed a split plot design with accession as main factor and isolate as secondary factor in 3 replicates. For the survey, there were a significant difference in disease incidence and severity. The district of Boukoumbe had the least disease incidence (74.44%) and severity (24.54%). The districts of Manta, Korontiere and Dipoli had the highest disease incidence (100%) and the district of Manta recorded the highest disease severity (53.08%). There were significant difference in disease incidence ($p = 0.004$) between the accessions of farmers during survey whereas all the accessions did not differ significantly from each other in respect to disease severity. After isolation, cultures observed were identified as *Curvularia* sp. For the experiment, 3 accessions were resistant, 6 were moderately resistant, 6 were moderately susceptible and 8 were susceptible. Moreover, a significant correlation between disease severity and seed weight were observed ($r = -0.81$ and $p = 2.2e^{-16}$), and seed weight loss due to the disease up to 45% was reported. In conclusion, Curvularia leaf spot is a constraint to fonio cultivation in the commune of Boukoumbe in the north Benin and resistant accessions exist and could be recommended to the farmers.

2 INTRODUCTION

White fonio (*Digitaria exilis* Stapf) is a neglected native crop within the West African sub-region (Bakasso et al., 2010). It grows fast and thrives on very poor soils extending from 8° to 14° N

latitude and from Senegal to Lake Chad (Cruz et al., 2011). It is cultivated from sea level up to 1500 m altitude and mainly in areas receiving annually 700 to 1000 mm rainfall, however in

critical rainfall deficiency conditions its limit is at the annual isohyet of 150 mm. The optimal growing temperature range is 25-30° C with approximately 12 h daylight (Haq and Ogbe, 1995). In Benin, fonio is produced in the communes of Boukombé, Cobly, Toucouthouan, Tanguiéta and Natitingou in the northwest of the country with the commune of Boukombé providing 74% of the national production (Dramé and Cruz, 2002; MAEP, 2010). It is cultivated under sudano guineen climate where rainfall ranges from 800 to 950 mm/year (Vodouhè *et al.*, 2003). Fonio is considered as the most nutritious cereals in Africa (Jideani, 1990) and can contribute to nutrition security as it contains all twenty amino acids in particular two essential ones (methionine and cysteine) (Vietmeyer *et al.*, 1996). It is the main food during traditional ceremonies (Adoukonou-Sagbadja *et al.*, 2006). Available statistics show that 631246 t of fonio are produced from 912721 ha by the world three largest producers Guinea, Nigeria and Mali. In Benin 1155 t of fonio were produced from 1868 ha (FAO Stat, 2017). Fonio yield depending on countries ranges between 200 and 900 Kg/ha (Vodouhè *et al.*, 2003). Fonio has a large diversity

3 MATERIALS AND METHODS

3.1 General survey: A disease survey was conducted in 19 sites of the commune of Boukoumbe in Benin during the sheaving period (July) in 2018. Two to three sites were surveyed

of accessions with different life cycle duration, which could play an important role in preventing food shortages (Blench, 2012; Gessain, 1989). Despite its importance, fonio production faces several challenges as poor soil fertility, use of low-yielding cultivars, inadequate weeding, pests and diseases, high harvest losses, laborious post-harvest management and low socio economic appraisal (FiBL, 2013). Although leaf spot have been reported on the crop caused by the fungi *Phyllachora sphearosperma*, *Helminthosporium* spp, *Curvularia* spp, *Puccinia cabuensis* and *Fusarium* spp. (Adoukonou-Sagbadja, 2010; Mamza *et al.*, 2013; Akanmu *et al.*, 2013), very little research work has been performed to confirm the causal pathogen. In addition, fonio cultivation relies only on traditional accessions, which are less productive, and their resistance to leaf spot are not well documented. However, a good management strategy of the leaf spot required selection of productive and resistant fonio accessions. Thus, the objectives of this study were to determine the distribution of the leaf spot of the white fonio in the commune of Boukoumbe, to identify its causal agent and to evaluate white fonio accessions for their resistance to the leaf spot.

per district within the 6 districts of the commune. The sites with at least 0.5 ha and 5km distance from each other were selected (Fig. 1).

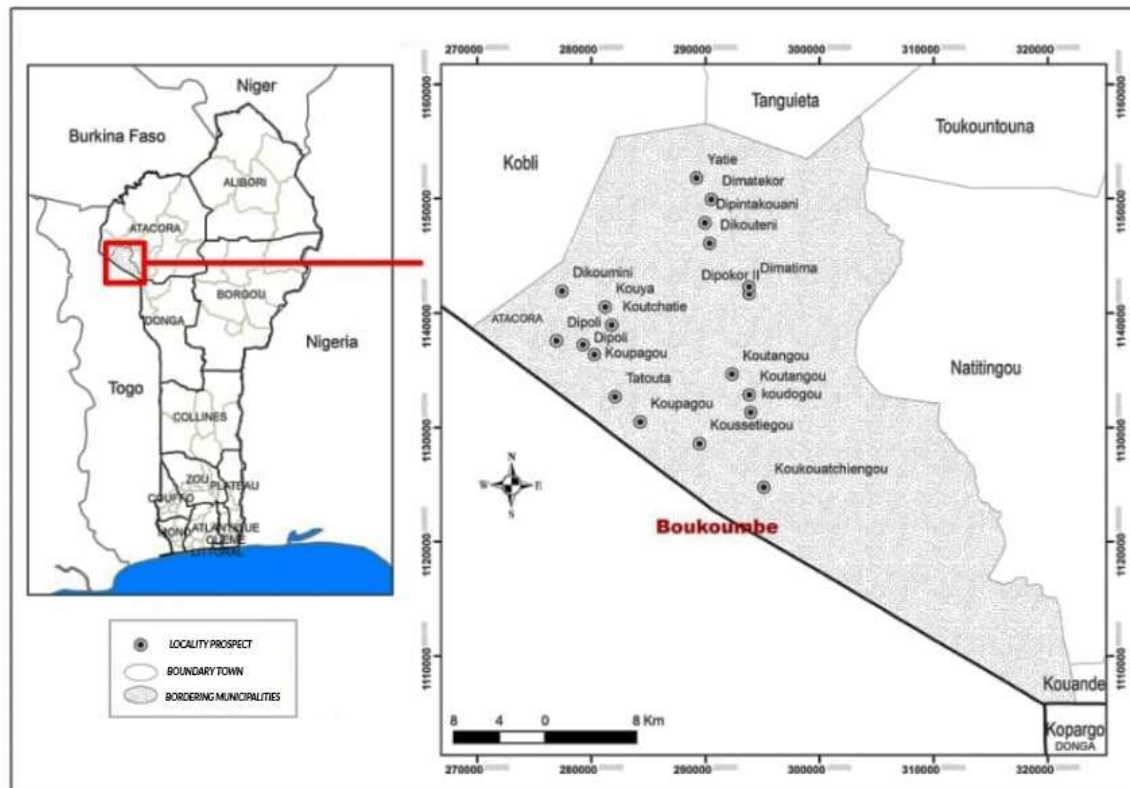


Fig. 1: White fonio production sites surveyed in the commune of Boukoubme

4 RESULTS

The commune of Boukoubme is located in the Northwest of Benin in the Department of Atacora. It's ranged between 10° and 10°40' N latitude and 0°754 and 1°30'E longitude in the Sudanian zone. The climate is Soudan-Guinean type and characterized by a rainy season from April to October and a dry season from November to March (Afrique Conseil, 2006). Disease symptoms with reddish brown spots or lesions occurs which may increase in size and become irregular. At each site, 30 plants were randomly selected on two diagonals through the field to evaluate the disease incidence. Ten of these plants were used to assessed index by scoring individual plants using a 1-9 scale described by Subrahmanyam *et al.* (1995) where 1= 0% of symptoms on leaves, 2= 1-5% of leaves infected (l.i.), 3= 6-10% of l.i., 4= 11-20% of l.i., 5= 21-30% of l.i., 6= 31-40% of l.i., 7= 41-60% of l.i., 8= 61-80% of l.i., 9= 81-100% of l.i. These assessments were used to calculate the disease severity as follow:

$$S = \frac{\sum n}{N \times 9} \times 100$$

with **S** = Disease severity, **n** = individual index, **N** = Total number of plant assessed, **9**: High index value.

These quantitative survey data were coupled with global position systems (GPS) and geographic information systems (GIS) technologies to map surveyed sites. Leaves with symptoms were collected and pathogens isolated.

4.2 Isolation and identification of *Curvularia* spp.: The disease leaves were cut and washed in thrice in sterile distilled water. The pieces were surface disinfected in a solution of 1% of sodium hypochlorite (NaOCl) for one minute and further rinsed in sterile distilled water and blotted dry. About 2mm² of leaf surface containing the disease and the healthy parts were cut and placed on Potato Dextrose Agar (PDA) supplemented with three drops of citric acid in Petri dishes. These dishes were then

incubated at 25°C for 72h and sub-cultures were done. The plates were incubated for 25°C for 7-10 days. Fungal structures are suspended in water and then in lactophenol on microscopic slide and observed under the microscope. Identification of the fungi was done using a key based on microscopic characteristics (nature of mycelium, morphology and formation of the conidiosporus, the arrangement of the conidia on the conidiosporus, the shape and size of the conidia, the type and number of conidia) (Watanabe, 1994).

4.3 Evaluation of white fonio accessions for resistance to *Curvularia* spp. experiment

4.3.1 Experimental design: Trials were conducted with twenty-three accessions coming from the seed stock at the Faculty of Agronomy (University of Parakou in North Benin) from July to October 2018. The experiment followed a split plot design with the fonio accession as main factor and the isolate as secondary factor. The system consisted of three replicates (blocks) subdivided into three sub-blocks established in a greenhouse at 21-32°C and 60-96%. Each sub-block was composed of twenty-three elementary plots (1 plot represented 2 pots 1.5dm³). The soil substrate was sterilized at 65°C for 72 h, then mixed with compost at the ratio of 3/4 (soil) and 1/4 commercial compost. No fertilizer was applied.

4.3.2 Inoculum preparation, inoculation and disease assessment: Conidia suspensions were prepared with sterilized distilled water and adjusted to 2 10⁴ conidia/ml using a hemacytometer. The prepared inoculum was applied to leaf surfaces with a hand sprayer after adding one drop of Tween 20. Plants sprayed with sterilized distilled water served as control. The inoculation was made from two virulent isolates of *Curvularia* spp. out of twenty-three accessions at the rate of three replicates per isolate. Two weeks after inoculation, disease index were evaluated on greenhouse plants by scoring individual plants using a 1-9 scale described by Subrahmanyam *et al.* (1995). Four disease index assessment were performed at two

weeks apart. The values from these assessments were used to calculate the disease severity per plot and then the area under severity curve as follow:

$$AUSPC = \sum_i [(S_i + S_{i-1}) * (t_i - t_{i-1})] / 2$$

with **t_i** = date of assessment, **S_i** = average of severity at t_i (Shaner and Finney, 1977; Jeger and Viljanen Rollinson, 2001).

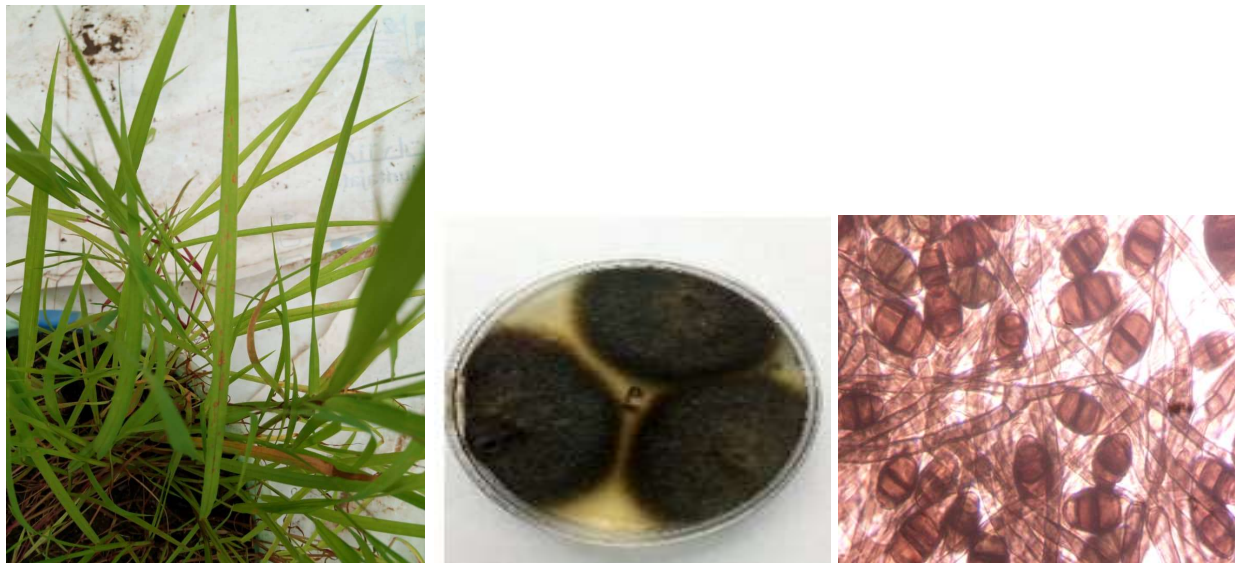
4.4 Statistical analysis: The disease incidence, severity and seed weight data (survey and greenhouse trial) which were log-transformed and subjected to analysis of variance (ANOVA) using the generalized linear model (GLM) procedure in R version 3.5.0 software (R Core Team 2018). Tukey test was performed to compare mean values of disease incidence, severity and seed weight. Values given in tables are original means with corresponding standard errors. Pearson correlation was analysed between disease severity and seed weight.

4.5 Distribution of *Curvularia* leaf spot and disease reaction of accessions of fonio in the commune of Boukoumbe: White fonio leaves collected during the survey showed symptoms with reddish brown spots or lesions occurs which may increase in size and become irregular (Fig. 2). The fungus isolated from diseased leaves was *Curvularia* sp. Mycelia were black presenting irregular borders and cottony growth and conidia were poroporous, subelliptical, cylindrical mainly 4-celled, dark brown and larger in central 2 cells (Figs. 3a, 3b). There were a significant difference in disease incidence and severity ($p < 2e^{-16}$). The district of Boukoumbe had the least disease incidence (74.44%) and severity (24.54%). The districts of Manta, Korontiere and Dipoli had the highest disease incidence (100%) followed by the districts of Natta (96.67%) and Tabota (95.83%). However, the district of Manta recorded the highest disease severity (53.08%) followed by the districts of Korontiere (44.93%), Dipoli (40.86%), Tabota (42.22%) and Natta (35.43%) (Table 1).

Table 1: Disease incidence and severity of *Curvularia* leaf spot of fonio in the commune of Boukoumbe

District	Incidence (%)	Severity (%)
Boukoumbe	74.44 ± 4.62 a	24.57 ± 1.17 a
Dipoli	100.00 ± 0.00 b	40.86 ± 1.16 c
Korontiere	100.00 ± 0.00 b	44.93 ± 1.15 c
Manta	100.00 ± 0.00 b	53.08 ± 1.69 d
Tabota	95.83 ± 1.83 b	42.22 ± 1.23 c
Natta	96.67 ± 1.90 b	35.43 ± 1.28 b
Average	94.56 ± 0.95	40.29 ± 0.64
Probability (P)	2e ⁻¹⁶	2e ⁻¹⁶
CV	0.24	0.38

Means with the same letter in column are not significantly different at 5% threshold for Turkey test

**Fig. 2:** Brown spots on leaf **Fig. 3:** (a) *Curvularia* sp. on PDA and (b) Conidia on microscope (1000 X)

Farmers sowed five accessions during our survey period (Ikantoni, Ipomoani, Iponi, Iporapia and Iprawan). There were significant differences in disease incidence ($p = 0.004$) between the accessions whereas all the accessions did not differ significantly from each other in respect to disease severity with accession Ipomoani having

the highest (45%). Accessions Ikantoni, Ipomoani and Iponi recorded high incidence and were at par with each other (100%) followed by accessions Iporapia and Iporawan having the least (95.33% and 91.11%, respectively) (Table 2).

Table 2: Disease reaction of different accessions of fonio to *Curvularia* leaf spot during the survey in the district of Boukoumbe

Accession	Incidence (%)	Sévérité (%)
Ikantoni	100.00 ± 0.00 b	38.51 ± 1.30 a
Ipomoani	100.00 ± 0.00 b	45.00 ± 1.43 a
Iponi	100.00 ± 0.00 b	36.67 ± 1.77 a
Iporapia	95.33 ± 1.72 ab	40.89 ± 1.12 a
Iporawan	91.11 ± 1.73 a	39.71 ± 1.08 a
Average	94.56 ± 0.95	40.29 ± 0.64
Probability (P)	0.00441 **	0.0635
CV	0.24	0.38

Means with the same letter in column are not significantly different at 5% threshold for Turkey test

4.6 Screening of white fonio accessions for resistance to *Curvularia* spp in semi-controlled conditions: There were significant differences in disease severity (AUSPC) of the two isolates (Bouk-S2 and Bouk-S3) and of some accessions. The disease severity ranged from 1,768 to 2,351 for the isolate Bouk-S3 and from 1,805 to 2,750 for the isolate Bouk-S2. The least disease severity was recorded on the accession AS4 for the isolate Bouk-S3 and on the accession AS3 for the isolate Bouk-S2. The highest disease severity were observed on accessions AS8, AS14, AS15 and AS21 for the isolate Bouk-S3 and AS2 and AS17 for the isolate Bouk-S2) (Table 3). Thus, the accessions were separated in four groups : accessions AS3, AS4 and AS22 were

resistant, accessions AS5, AS9, AS10, AS11, AS12 and AS23 were moderately resistant, accessions AS6, AS7, AS16, AS18, AS19 and AS20 were moderately susceptible, accessions AS1, AS2, AS8, AS13, AS14, AS15, AS17 and AS21 were susceptible (Fig. 4). Analysis of the Pearson correlation between disease severity and seed weight showed a significant negative correlation ($r = -0.81$ and $p = 2.2e^{-16}$). Analysis of the different accessions showed that the two isolates reduced the weights compared to the control except for accession AS17. Weight losses ranged from 4.65 to 41.82% for the isolate Bouk-S3 and from 0.39 to 45.18% for the isolate Bouk-S2 (Table 4).

**Table 3:** Disease severity (AUSPC) of two isolates of *Curvularia* sp. on 23 white fonio accessions in semi-controlled conditions

<i>Accession local Name</i>	<i>Accession number</i>	<i>Isolate Bouk-S3</i>	<i>Isolate Bouk-S2</i>	<i>Average AUSPC</i>	<i>F</i>	<i>P</i>
<i>Yoro</i>	AS1	2240.74 ± 96.66 ab A	2592.59 ± 137.10 ce A	2416.67 ± 116.88 c	4.399	0.0522
<i>Iporni</i>	AS2	2166.66 ± 166.08 ab A	2685.18 ± 129.38 e B	2425.93 ± 147.73 c	6.066	0.0255
<i>Iporda</i>	AS3	2138.88 ± 112.83 ab A	1796.29 ± 66.76 a B	1967.59 ± 89.81 a	6.828	0.0188
<i>Affésiamon</i>	AS4	1768.51 ± 38.73 a A	2287.03 ± 139.65 ae B	2027.78 ± 89.19 ab	12.8	0.00251
<i>Lamba</i>	AS5	2305.55 ± 63.64 ab A	2027.77 ± 53.79 abcd B	2166.67 ± 58.72 ac	11.11	0.00421
<i>Not identified</i>	AS6	2055.55 ± 109.36 ab A	2379.63 ± 157.20 ae A	2217.59 ± 133.28 bc	2.864	0.11
<i>Ipormondé</i>	AS7	2287.03 ± 134.73 ab A	2287.03 ± 181.14 ae A	2287.04 ± 157.94 bc	0	1
<i>Ikantoni</i>	AS8	2351.85 ± 88.69 b A	2361.11 ± 121.87 ae A	2356.48 ± 105.28 cd	0.004	0.952
<i>Iporgnirmé</i>	AS9	2268.51 ± 144.85 ab A	1861.11 ± 70.81 ab B	2064.81 ± 107.84 ab	6.384	0.0224
<i>Iporawan</i>	AS10	2175.92 ± 110.62 ab A	1981.48 ± 45.59 abc A	2078.70 ± 78.11 ab	2.641	0.124
<i>Iporapia</i>	AS11	2129.63 ± 86.85 ab A	2009.25 ± 113.21 abc A	2069.44 ± 100.04 ab	0.712	0.411
<i>Iporapia</i>	AS12	1935.18 ± 116.84 ab A	2314.81 ± 161.83 ae A	2125 ± 139.34 ab	3.617	0.0754
<i>Iporapia</i>	AS13	2277.77 ± 114.53 ab A	2388.88 ± 205.06 ae A	2333.33 ± 159.80 cd	0.224	0.643
<i>Ipomonpiété</i>	AS14	2324.07 ± 100.58 b A	2675.92 ± 123.79 de B	2500 ± 112.1876 c	4.866	0.0424
<i>Tignonpété</i>	AS15	2314.81 ± 45.59 b A	2370.37 ± 131.10 ae A	2342.59 ± 88.35 cd	0.16	0.694
<i>Iporawan</i>	AS16	2268.51 ± 145.52 ab A	2175.92 ± 133.53 ae A	2222.22 ± 139.53 bc	0.22	0.646
<i>Yoro</i>	AS17	2259.25 ± 80.32 ab A	2750.00 ± 103.00 e B	2504.63 ± 91.66 c	14.12	0.00172
<i>Tignonpété</i>	AS18	2287.03 ± 78.70 ab A	2268.51 ± 131.59 ae A	2277.78 ± 105.15 bc	0.015	0.905
<i>Iporni</i>	AS19	2203.70 ± 114.62 ab A	2231.48 ± 150.72 ae A	2217.59 ± 132.68 bc	0.022	0.885
<i>Iporawan</i>	AS20	2101.85 ± 79.51 ab A	2509.25 ± 81.51 be B	2305.55 ± 80.51 cd	12.8	0.00251
<i>Iporapia</i>	AS21	2333.33 ± 70.81 b A	2379.63 ± 149.01 ae A	2356.48 ± 109.9 cd	0.079	0.783
<i>Tignonpété</i>	AS22	2000.00 ± 105.77 ab A	1805.55 ± 79.78 a	1902.78 ± 92.78 a	2.154	0.162
<i>Iporni</i>	AS23	2092.59 ± 77.88 ab A	2157.40 ± 74.07 ae A	2125 ± 75.98 ab	0.364	0.555
	Average AUSPC	2186.39 ± 22.73	2273.75 ± 30.97			
	F	1.939	4.667			
	P	0.00974	0.00000000175			

Means with the same letter in column and in row are not significantly different at 5% threshold for Turkey test

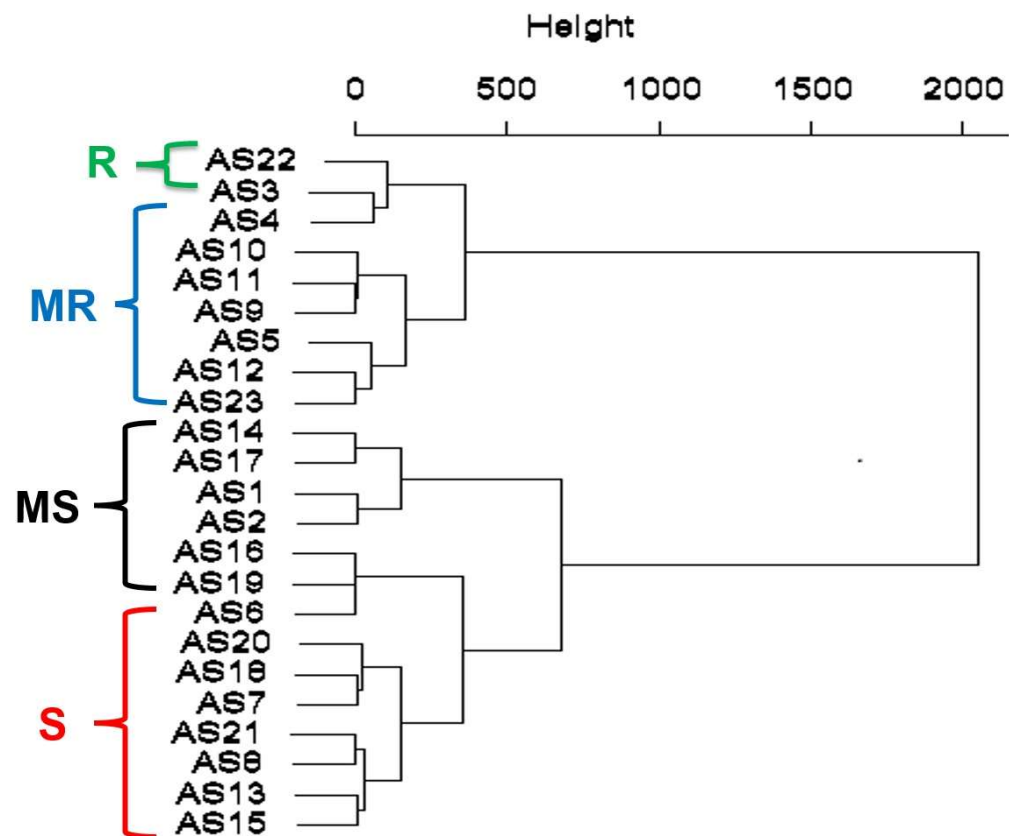


Fig. 4: Dendrogram showing the disease severity (AUSPC) reaction groups of white fonio accessions after inoculation with two isolates of *Curvularia* sp. in semi-controlled conditions

**Table 4:** Seed weight (g) and seed weight loss (%) of 23 white fonio accessions after inoculation with two *Curvularia* sp. isolates

<i>Accession local Name</i>	<i>Accession number</i>	<i>Seed weight (g) Control</i>	<i>Seed weight (g) for inoculated plants by isolate Bouk-S3</i>	<i>Seed weight loss (%) for inoculated plants by Bouk-S3</i>	<i>Seed weight (g) for inoculated plants by isolate Bouk-S2</i>	<i>Seed weight loss (%) for inoculated plants by Bouk-S2</i>
<i>Yoro</i>	AS1	11.80 ± 0.99 abA	8.80 ± 0.92 aB	-25.42	9.24 ± 0.85 bB	-21.69
<i>Iporni</i>	AS2	12.36 ± 0.56 bA	7.45 ± 0.21 aB	-39.72	7.84 ± 0.90 aB	-36.57
<i>Iporda</i>	AS3	12.00 ± 0.6 abA	9.33 ± 1.15 aB	-22.25	7.96 ± 0.07 aB	-33.67
<i>Affèsiamon</i>	AS4	12.46 ± 0.49 bA	7.79 ± 0.69 aB	-37.48	6.83 ± 0.30 aB	-45.18
<i>Lamba</i>	AS5	12.36 ± 0.84 bA	11.18 ± 1.44 aB	-9.55	9.59 ± 0.29 bB	-22.41
<i>Non Identifié</i>	AS6	10.60 ± 0.86 aA	8.37 ± 0.69 aB	-21.04	8.34 ± 1.06 aB	-21.32
<i>Ipormondé</i>	AS7	12.12 ± 0.51 abA	9.14 ± 0.64 aB	-24.59	9.45 ± 0.27 bB	-22.03
<i>Ikantoni</i>	AS8	9.56 ± 0.96 aA	8.37 ± 1.01 aB	-12.45	6.52 ± 0.92 aB	-31.80
<i>Iporgnirmè</i>	AS9	12.28 ± 0.92 bA	9.25 ± 0.48 aB	-24.67	9.59 ± 1.36 bB	-21.91
<i>Iporawan</i>	AS10	12.56 ± 0.46 bA	8.80 ± 0.23 aB	-29.94	8.93 ± 0.68 abB	-28.90
<i>Iporapia</i>	AS11	12.36 ± 0.74 bA	10.56 ± 0.72 aB	-14.56	9.07 ± 0.75 bB	-26.62
<i>Iporapia</i>	AS12	12.28 ± 0.76 bA	9.26 ± 0.25 aB	-24.59	8.86 ± 1.27 abB	-27.85
<i>Iporapia</i>	AS13	12.00 ± 0.75 abA	8.92 ± 1.11 aB	-25.67	9.59 ± 0.41 bB	-20.08
<i>Ipomonpiété</i>	AS14	12.28 ± 0.36 bA	9.37 ± 0.15 aB	-23.70	9.98 ± 0.67 bB	-18.73
<i>Tignonpété</i>	AS15	12.48 ± 0.81 bA	8.91 ± 0.96 aB	-28.61	8.36 ± 0.66 aB	-33.01
<i>Iporawan</i>	AS16	12.40 ± 1.2 bA	9.76 ± 1.48 aB	-21.29	9.72 ± 0.93 bB	-21.61
<i>Yoro</i>	AS17	10.32 ± 0.73 aA	10.80 ± 0.79 aB	4.65	10.36 ± 0.67 cB	0.39
<i>Tignonpété</i>	AS18	12.48 ± 0.79 bA	9.37 ± 1.22 aB	-24.92	9.01 ± 0.27 bB	-27.80
<i>Iporni</i>	AS19	12.28 ± 0.51 bA	8.67 ± 0.84 aB	-29.40	10.73 ± 0.19 cB	-12.62
<i>Iporawan</i>	AS20	12.20 ± 1.55 abA	10.06 ± 1.68 aB	-17.54	9.73 ± 1.43 bB	-20.25
<i>Iporapia</i>	AS21	12.10 ± 0.22 abA	7.04 ± 0.11 aB	-41.82	8.87 ± 0.33 abB	-26.69
<i>Tignonpété</i>	AS22	12.58 ± 0.69 bA	10.19 ± 0.78 aB	-19.00	10.32 ± 0.59 cB	-17.97
<i>Iporni</i>	AS23	12.38 ± 0.63 bA	7.90 ± 0.41B	-36.19	8.04 ± 0.84 aB	-35.06
<i>F</i>		1.923	1.346		1.877	
<i>P</i>		0.013	0.195		0.036	

Means with the same letter in column are not significantly different at 5% threshold for Turkey test

5 DISCUSSION

This study highlights the importance of the *Curvularia* leaf spot in the commune of Boukoumbe. The disease incidence and severity differed across sites. The highest incidence and severity values at Dipoli, Korontiere and Manta showed that these districts are more vulnerable to fonio leaf spot. This may be due to environmental conditions (temperature, relative humidity and level of soil fertility). This finding confirmed the report of Manza *et al.* (2013) in Riyom, Plateau State of Nigeria where highest disease incidence and severity of *Curvularia* leaf spot occurred on some sites. In addition, Ou (1985) and Percich *et al.* (1997) stated that these factors influence the expression of the disease aggressiveness. Aouali and Douici-Khalfi (2013) reported that disease severity depends on the climatic conditions, the variety and the stage of development of the plant at the time of the attack. Moreover, the production system of fonio in the commune of Boukoumbe based generally on monoculture without crop rotation on the same land and on the none elimination of weeds or crop residues that are potential sources of inoculum. The lowest incidence value was observed on the accession Ipordawan while the highest was observed on the accessions Ikantoni, Ipormi and Ipomoani. This could be explained by the duration of the cycle of these different accessions. In fact, Sekloka *et al.* (2016) stated that Ikantoni and Iporni belong respectively to the short cycle, extra early (2.5 to 3 months) and late cycle (5 months) accession groups unlike Ipordawan has a duration cycle from 3.5 to 4

months. In addition, Aouali and Douici-Khalfi (2013) who noticed that wheat leaf spot severity depend on several parameters including the development stage of the plant during the infection process have done the same report. All management strategies for fonio disease control required selection of productive and resistant accessions. Thus, in this study twenty-three accessions of fonio tested in the greenhouse to *Curvularia* leaf spot reacted differently to the disease. Among them, 3 accessions were resistant, 6 were moderately resistant, 6 were moderately susceptible and 8 were susceptible. The observed diversity in disease reactions of accessions were obtained by Manza *et al.* (2013) with sixteen accessions of *D. iburua* in Nigeria. The results corroborate those obtained by Oubda (2014) in Burkina Faso and Ramathani (2010) in Uganda who reported variability between some varieties of maize and sorghum tested against leaf spot. This variability could be due to differential responses of the resistance genes or not (Saifuila, 1994; Datnoff and Lentini, 1994; Rai *et al.*, 2004). The polymorphic SSRs currently available for white fonio (Clément and Leblanc, 1984; Barnaud *et al.* 2012; Olodo *et al.*, 2018) could be used for accurate genetic studies in relation to *Curvularia* leaf spot. Also, a significant negative correlation between disease severity and seed weight, and seed weight losses up to 45% were observed showing that *Curvularia* leaf spot is an important disease, which negatively affect fonio production.

6 ACKNOWLEDGEMENTS

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7 REFERENCES

- Adoukonou-Sagbadja H. 2010. Genetic characterization of traditional fonio millets (*Digitaria exilis*, *Digitaria iburua* Stapf) landraces from West-Africa: implications for conservation and breeding. PhD thesis. Justus-Liebig University, Giessen, Germany, 119p.
- Adoukonou-Sagbadja H, Dansi A, Vodouhe R. and Akpagana, K. 2006. Indigenous knowledge and traditional conservation of Fonio millet (*Digitaria exilis* Stapf, *Digitaria iburua* Stapf) in Togo. Biodiversity and Conservation 15:2379–2395. doi:10.1007/s10531-004-2938-3.

- Afrique Conseil 2006. Monographie de la Commune de Boukombé. Programme d'appui au démarrage des Communes, Mission de décentralisation. République du Bénin. Rapport de consultation. 61 p.
- Akanmu A.O., Abiala M.A. and Odebode A.C. 2013. Pathogenic Effect of Soil borne Fusarium Species on the Growth of Millet Seedlings. World Journal of Agricultural Sciences 9 (1): 60-68. DOI: 10.5829/idosi.wjas.2013.9.1.1721.
- Aouali S. et Douici-Khalfi A. 2013 : Recueil des principales maladies fongiques des céréales en Algérie : symptômes, développement et moyens de lutte. ITGC. 8-36.
- Bakasso Y, Barry M. B., Bezançon G., Cruz J.-F., Noyer J.-L. and Pham J.-L. 2010. De la connaissance à la valorisation du fonio Atelier international, 9-11 décembre à Niamey (Niger). 1p.
- Barnaud A., Vignes H., Risterucci A.-M, Noyer J.-L., Pham J.-L., Blay C and Billot C. 2012. Development of nuclear microsatellite markers for the fonio, *Digitaria exilis* (Poaceae), an understudied West African cereal. American Journal of Botany 99: 105–107.
- Blench R.M. 2012. Vernacular Names for African Millets and Other Minor Cereals and Their Significance for Agricultural History. Archaeological and Anthropological Sciences, October. DOI:10.1007/s12520-012-0104-5.
- Clément J. et Leblanc J.M. 1984. Prospection des *Digitaria exilis* (Fonio) en Afrique de l'Ouest. Marseille, France : ORSTOM.
- Cruz J.F., Béavogui F. et Drame D. 2011. Le fonio, une céréale africaine. 175p.
- Datnoff L.E. and Lentini R. S. 1994. Brown Spot in Florida Rice. Plant Pathology Department. Institute of Food and Agricultural Sciences. University of Florida. Publication May 1994, 128 p.
- Dramé D. et Cruz J. 2002. Rapport de Mission au Bénin et au Sénégal. Amélioration des technologies post-récolte du fonio (CORAF). 21 p.
- FiBL 2013: African Organic Agriculture Training Manual. Version 2.0 July 2013. Edited by Gilles Weidman and Lukas Kilcher. Research Institute of Organic Agriculture FiBL, Frick
- Gessain M. 1989. Youkounkoun Revisited. Journal des Africanistes 59:163–83. DOI:10.3406/jafr.1989.2281.
- Haq N. and Ogbe D. F. 1995. Fonio (*Digitaria exilis* and *D. iburua*). In: Williams, J.T. (Editor). Cereals and pseudo cereals. Chapman and Hall, London, United Kingdom. pp. 225–245.
- Jeger M.J. and Viljanen-Rollinson L.H. 2001. The use of the area under the disease progress curve (AUDPC) to assess quantitative disease resistance in crop cultivars. Theoretical and Applied Genetics 102 (1):32-40.
- Jideani I.A. 1990. Acha, (*Digitaria exilis* Stapf.), the neglected cereal. Agriculture International 42:132- 134.
- MAEP 2010. Ministère de l'Agriculture de l'Elevage et de la pêche /CeRPA/Service Statistique. <http://www.countrystat.org/ben/cont/pxwebquery/ma/053spd010/fr/vType/quick>.
- Mamza W.S., Zarafi A.B, Dachi S.N., Bright E.O. and Ukuwngwu M.N. 2013. Incidence and severity of *Curvularia* leaf spot on *Digitaria iburua* (fonio) in Riyom, Plateau State, Nigeria. P 88. In: Hall, R.A., P. Rudebjer, S. Padulosi. (eds.) 2013. 3rd International Conference on: Neglected and Underutilized Species (NUS): for a Food-Secure Africa. Accra, Ghana, 25-27 September 2013. Book of Abstracts. Bioversity International, Rome, Italy.
- Olodo K.F., Gueye M.C., Calatayud C., Diop B.M., Kane N.A., Ngom A., Ntui V.O., Barreto M-M. S., Uyoh E.A., Abraham S., Vigouroux Y., Billot C. and Barnaud A. 2018. EST-SSR development for *Digitaria exilis* and its relatives *D. iburua* and *D. longiflora* from transcriptome sequences. Plant Genetic Resources; 1–5.

- Ou S.H. 1985. Rice Diseases. 2nd ed. Commonwealth Mycological Institute, ISBN 0 85 198 545 9, Ferry Lane, Kew, Surrey, England. 380 p.
- Oubda W.Y.C. 2014. Evaluation de lignées stables obtenues dans des variétés de maïs de cycles intermédiaire, précoce et extra-précoce vulgarisées au Burkina Faso. Mémoire de fin de cycle pour l'obtention du diplôme de Master en Production Végétale. Institut du Développement Rural. Université Polytechnique de Bobo-Dioulasso. Burkina-Faso. 100p.
- Percich J.A., Nyvall R. F. and Malvick, D. K. 1997. Interaction of temperature and moisture on infection of wild rice by *Bipolaris oryzae* in the growth chamber. *Plant Disease* 81:1193-1195.
- Rai B., Jha, M. M. and Jha, A. C. 2004. Screening of resistant sources of rice against brown spot disease. *Annals of Biology*. 20 (2):189-193.
- Ramathani I. 2010. Characterisation of Turcicum leaf blight epidemics and pathogen populations in the *Exserohilum turcicum* – sorghum pathosystem of Uganda. A thesis submitted to the school of graduate studies in partial fulfillment of the requirements for the award of a Master of Science degree in crop science of Makerere University. pp 1-138.
- R Core Team. 2018. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria.
- Saifuila M. 1994. Field screening of rice genotypes to brown spot and leaf scald disease. *Agricultural science digest*: 14 (1) 68–70.
- Shaner G. and Finney R.E. 1977. The effect of nitrogen fertilization on the expression of slow-mildewing resistance in knows wheat. *Phytopathol.*, 67, 1051-1056.
- Sekloka E., Kanlindogbe C., Biauou S.S.H., Adoukonou-Sagbadja H., Kora A., Motouama F. T., Seidou M., Zinsou V.A., Afouda L. and Baba-Moussa L. 2016. Agro-morphological characterization of fonio millet accessions (*Digitaria exilis* Stapf.) collected from Boukoumbé, Northwest of Benin. *Journal of Plant Breeding and Crop Science*. 8 (10):211-222.
- Subrahmanyam P., McDonald D., Walliyar F., Raddy L.J., Nigam S.N., Gibbons R.W., Rammanatha R.V., Singh A.K., Pande S., Reddy P.M. and Subba Rao P.V. 1995. Screening methods and sources of resistance to rust and late leaf spot of groundnut. *Bull.* 47. ICRISAT, Patancheru, India, 20p.
- Vietmeyer N.D., Borlaugh N.E., Axtell J., Burton G.W., Harlan J.R. and Rachie K.O. 1996: Fonio (Acha). In: *BOSTID Publication. Lost crop in Africa*, In: *BOSTID Publication. Lost crop in Africa*. Pp 59–75.
- Vodouhè S.R., Zannou A. et Achigan Dako E.G. 2003. Actes du premier atelier sur la diversité génétique du fonio (*Digitaria exilis* Stapf.) en Afrique de l'Ouest. Conakry, Guinée, 4–6 août 1998. IPGRI, Rome, Italy. 73 p.
- Watanabe T. 1994. Pictorial of Atlas of Soil and Seed Fungi: Morphologies of cultured fungi and key to species. CRC Press, Inc. Lewis Publishers. 411p.